

ikhlas jenie turnitin 1

by Ikhlas Jenie

Submission date: 24-Sep-2019 12:30PM (UTC+0700)

Submission ID: 1178898251

File name: 4577-7053-1-PB-Cytoprotective_effect.pdf (267.54K)

Word count: 2161

Character count: 12741

Cytoprotective effect of tangeretin in hydrogen peroxide-induced oxidative stress on human umbilical vein endothelial cells (HUVECs)

Riris Istighfari Jenie¹, Irfan Muris Setiawan¹, Ikhlas Muhammad Jenie², Haruma Anggraini Muflikhasari³

¹Faculty of Pharmacy, Universitas Gadjah Mada, Yogyakarta, Indonesia

²Faculty of Medicine and Health Sciences, Universitas Muhammadiyah Yogyakarta, Yogyakarta, Indonesia

³Faculty of Biology, Universitas Gadjah Mada, Yogyakarta, Indonesia

Email: riris_jenie@ugm.ac.id

ABSTRAK

Cekaman oksidatif diketahui merupakan pemicu terhadap kerusakan jaringan vaskuler yang kemudian mengakibatkan timbulnya penyakit hipertensi, aterosklerosis, dan gagal jantung. Senyawa polimetoksilflavon yang terkandung dalam buah sitrus seperti tangeretin dan nobiletin telah diketahui memiliki aktivitas biologik, antara lain menurunkan kadar kolesterol dan sebagai antiinflamasi. Mekanisme penting yang memperantarai aterosklerosis adalah terjadinya cekaman oksidatif, yang menyebabkan disfungsi sel endotel. Penelitian ini bertujuan untuk mengetahui efek sitoprotektif tangeretin terhadap cekaman oksidatif pada sel HUVEC yang diinduksi dengan hidrogen peroksida. Viabilitas sel ditentukan melalui metode MTT. Sel diberi perlakuan seri konsentrasi tangeretin (12,5, 100, and 200 μM) selama 24 jam sebelum diberi perlakuan 600 μM H_2O_2 selama 4 jam. Analisis viabilitas sel menunjukkan bahwa tangeretin pada konsentrasi rendah, 12,5 μM , tangeretin menghambat sitotoksitas yang diinduksi oleh H_2O_2 dan bahkan menginduksi proliferasi sel. Namun demikian, pada konsentrasi tinggi, tangeretin tidak mampu menghambat sitotoksitas H_2O_2 dan justru menginduksi sitotoksitas yang lebih tinggi daripada H_2O_2 . Berdasarkan hasil ini, efek sitoprotektif tangeretin terhadap stress oksidatif memerlukan optimasi terhadap konsentrasi yang diaplikasikan.

Kata kunci: Tangeretin, cekaman oksidatif, sel endotel vena umbilical manusia (HUVEC), hidrogen peroksida.

INTRODUCTION

Reactive oxygen species (ROS) is a potent pathogenic mechanism in atherosclerosis, a major player in the initiation and progression of cardiovascular dysfunction associated with diseases such as hypertension, atherosclerosis, diabetes mellitus, and chronic heart failure (Taniyama and Griending, 2003). Oxidative stress, defined as

ABSTRACT

Oxydative stress is known as a major trigger in cardiovascular dysfunction that leads to hypertension, atherosclerosis, and chronic heart failure. Polymethoxyflavone such as tangeretin and nobiletin has been reported to have biological activity including reducing cholesterol level and anti inflammation activity. The critical point that mediates atherosclerosis is the formation of stress oxidative which causes endothelial cells dysfunction. This study was conducted to explore the effect of tangeretin in protecting H_2O_2 -induced oxidative stress on HUVEC. Cell viability was determined by using MTT assay. Cells were treated with different concentrations of tangeretin (12.5, 100, and 200 μM) for 24h before exposed to 600 μM H_2O_2 for 4h. Cells viability analyses showed that at low concentration (12.5 μM), tangeretin inhibited H_2O_2 -induced cytotoxicity. Furthermore, it induced cells proliferation. However, at high concentration, it failed to protect HUVEC from H_2O_2 -induced cytotoxicity and increased the cytotoxicity of H_2O_2 . Based on this result, tangeretin in low concentration has a potential to reduce oxidative stress on HUVECs.

Keywords: Tangeretin, oxidative stress, human umbilical vein endothelial cells (HUVECs), hydrogen peroxide.

imbalance of pro-oxidants and antioxidants in the organism (Taniyama and Griending, 2003). The ROS underlies redox chemistry involving reduction-oxidation reactions as formation of the univalent reduction of oxygen, i.e. superoxide (O_2^-) (Stocker and Koeney, 2004). Although superoxide itself has injury effects on vascular function, it is also play

major in producing other reactive species by several enzyme systems (Taniyama and Griendling, 2003; Stocker and Keaney, 2004). Dismutation of superoxide by superoxide dismutase enzyme (SOD) will produce the more stable ROS, i.e. hydrogen peroxide (H_2O_2), which is then converted enzymatically into H_2O by catalase and glutathione peroxidase (GPx) (Taniyama and Griendling, 2003). Under normal circumstances the levels of ROS are low enough to be removed by the natural defense systems in the body. Several antioxidant enzymes in the cells may convert the stable ROS into non-reactive form. Therefore, it inhibits the formation of more reactive ROS (Stocker and Keaney, 2004). Thus, homeostasis between production of ROS and antioxidant plays major role in the prevention of oxidative stress.

Oxidative stress is one of potent pathogenic mechanism in atherosclerosis (Hansson, 2005) by stimulating cellular injury, mitochondrial dysfunction, and apoptosis (Madamanchi and Runge, 2007). One possible mechanism by which oxidative stress may trigger cellular toxicity in cell is by inducing the release of cytochrome c into the cytosol, which in turn triggers the mitochondria apoptotic pathway (Wen et al., 2013). Atherosclerosis is a chronic inflammatory response to high serum cholesterol levels, leading to plaque formation and the hardening of arteries. Cumulatively, these effects increase an individual risk of stroke, myocardial infarction or additional systemic complications (Singhet et al., 2002). The progressive nature of this disease can remain undetected for many years (Hansson, 2005). Atherosclerosis induce endothelial dysfunction, that functions as an impermeable barrier whose integrity plays important roles in prohibiting leukocyte adhesion, reducing inflammation, and supporting the vasculature that release paracrine signaling peptides to regulate vascular tone during hemodynamic stress and oxidative stress (Davidson and Duchon, 2007). Therefore, it is need to develop new compound affecting multiple target like oxidative stress and mitochondria may afford better protection against atherosclerosis.

In natural source, flavonoids have been known as a potent antioxidant. Epidemiological studies have reported that diets rich in fruits, vegetables, and tea might reduce risk of chronic diseases such as cancer and cardiovascular diseases (Mulvihiland Huff, 2012). Polymethoxy-flavones (PMF) compounds in orange peel are one kind of flavonoids that was reported to have the ability in lowering-cholesterol levels (Kurowska et al., 2004). Previous report showed that water extract of sweet orange peel inhibited the cytotoxic effect of t-BHP on HepG2 cells. The phenolic bioactive compounds in the extract, including tangeretin (Fig.1), may responsible on that effect (Chen et al., 2012).

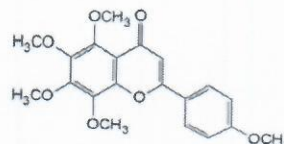


Figure 1. The molecule structure of tangeretin

To investigate whether tangeretin protected endothelial cells from oxidative stress, we used hydrogen peroxide (H_2O_2) to induce oxidative stress in human umbilical vein endothelial cells (HUVECs). In previous study (Wen et al., 2013), treatment with H_2O_2 in endothelial cell culture was reported to stimulate ROS concentration, decrease antioxidant enzymes, increase mitochondrial dis-function, and increase product of lipid peroxidation enzyme.

This study elucidates the potency of tangeretin to protect endothelial cells against oxidative stress and endothelia dis-function. Thus, it is potential to be developed to prevent atherosclerosis.

MATERIALS AND METHOD

Isolation and Cell culture

Human umbilical vein cells (HUVECs) are isolated from the vein of the umbilical cord by 1 mg/mL collagenase (kindly given by Prof. Hiroshi Itoh) treatment (Ethical clearance from Medical and

Health Research Ethics Committee (MHREC) Faculty of Medicine Universitas Gadjah Mada – Dr. Sardjito General Hospital No. KE/FK/619/EC). Cells seeded on gelatin coated dish and cultured in monolayers at 37°C in a 5% CO₂ atmosphere in Dulbecco modified Eagles medium (DMEM low glucose) (Gibco) supplemented with 10% heat inactivated 10% fetal bovine serum (FBS) (Sigma); 1% penicillin/streptomycin (Sigma); 0.5% fungizon (Gibco); 1 ng/mL vascular endothelial growth factor (VEGF) (Sigma); and 30 µg/mL endothelial cell growth factor (ECGF) (Sigma).

Cytotoxicity assay

Cell viability was measured by the MTT assay (Sigma), which measures the mitochondrial reduction MTT to formazan. HUVECs, 70-80% confluence, in 96-well plates were cultured in 10% FBS-DMEM containing 1% pen-strep at 37°C in 5% CO₂ atmosphere for 24 h. Cells were treated a series of concentrations of tangeretin (Sigma) (12.5; 100; and 200 µM) for 24h before exposed to 600 µM H₂O₂ (Merck) for 4h. Following H₂O₂ treatment, 0.5 mg/mL MTT was added to each well and cultures were incubated for 4 h at 37°C. The dark blue formazan crystals formed in intact cells were solved in 10% sodium dodecyl sulfate (SDS). The purple colour of the solution was measured by ELISA reader (BioRad) λ 550 nm and analyzed. Cell viability was calculated as follows:

Cell viability (%) = (absorbance of each treated group: absorbance of control) x 100%.

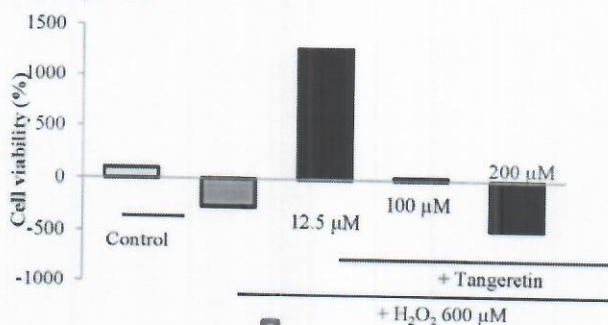


Figure 2. Profile the effect of tangeretin on Human umbilical vein endothelial cells (HUVECs) against hydrogen peroxide (H₂O₂) treatment. HUVECs were treated with different concentrations of tangeretin (12.5; 100; and 200 µM) for 24h, then exposed to 600 µM H₂O₂ for 4h. Following H₂O₂ treatment, the viability of the cells were then measured using MTT solution and as described in the method.

RESULTS AND DISCUSSION

The bioactive compounds contained in orange peel have been suggested to contribute in the effect of water extract of sweet orange peel in protecting HepG2 cells from oxidative stress induced by t-BHP (Chen et al., 2012). Tangeretin, together with nobiletin, is one of the polymethoxylated flavones that are abundant in the orange peel. This study explored the potency of tangeretin in protecting endothelial cells from oxidative stress by measuring cells viability using MTT.

We determined the effect of tangeretin in protecting endothelial cells by pre-treated the cells with different concentrations of tangeretin (12.5; 100; and 200 µM) for 24h, before exposing it to 600 µM H₂O₂ for 4h. As shown in Fig.2, treatment of H₂O₂ alone reduced cells viability much smaller than control cells. On the contrary, cells pre-treated with 12.5 µM tangeretin were not affected by H₂O₂ treatment and grow higher than control cells. However, high concentration of tangeretin, 100 and 200 µM, were failed to protect the cells from H₂O₂ treatment. These results showed that H₂O₂ (600 µM) was cytotoxic on HUVECs, and tangeretin had different effect depending on its concentration. Pre-treatment with low concentration of tangeretin preserved endothelial cell viability, but high concentration of tangeretin did not.

Moreover, low concentration tangeretin may have a proliferative effect for the endothelial cells because at this concentration, cells viability were higher than control, although it has been exposed to cytotoxic agent, H_2O_2 . From the present result (Fig.2), it is suggested that low concentration of tangeretin may protect cells from cytotoxicity caused by oxidative stress. However, high concentration induced cytotoxicity. The potential of tangeretin in reducing oxidative stress may involve several mechanisms such as reducing ROS and lipid peroxidation products, and increasing catalase enzyme activities. Tangeretin was reported to limit endogenous ROS production but it did not scavenge DPPH radical (Yoon et al., 2011). Wen et al. (2013) showed that H_2S inhibited H_2O_2 mediated mitochondrial dysfunction in human endothelial cells by preserving antioxidant defenses.

Therefore, it is important to further examine the antioxidant system and mitochondrial integrity, including analysis of ROS and lipid peroxidation products; endogen antioxidant enzyme activities; and expression of cytochrome c. This paper demonstrates the potential of tangeretin as a cytoprotective agent.

CONCLUSION

In conclusion, tangeretin in low concentration has a potential to reduce oxidative stress on HUVECs. However, high concentration of tangeretin cause cytotoxicity, therefore further experiments are needed to optimize its concentration as cytoprotective.

ACKNOWLEDGEMENT

1. Balaibangkes Kemenkes RI for the research grant from its program, RISBIN IPTEKDOK 2014.
2. Dokter Dwi Haryadi Sp. OG, for the umbilical cord samples.
3. Prof. Hiroshi Itoh from Nara Institute Science and Technology (NAIST) Japan for the collagenase.

REFERENCES

- Taniyama, Y. and Griendling, K.K. 2003. Reactive oxygen species in the vasculature: molecular and cellular mechanism. *Hypertension* 42: 1075-81.
- Stocker, R. and Keaney, J.F. Jr. 2004. Role of oxidative modifications in Atherosclerosis. *Physiol Rev* 84: 1381-1478.
- Hansson, G.K. 2005. Mechanisms of disease: Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* 352: 1685-1695+1626.
- Singh R.B., Mengi S.A., Xu Y-J, Arneja A.S., and Dhalla N.S. 2002. Pathogenesis of atherosclerosis-A multifactorial process. *ExpClinCardiol*. 7: 40-53.
- Madamanchi, N.R. and Runge, M.S. 2007. Mitochondrial Dysfunction in Atherosclerosis. *Circ Res*.100: 460-473.
- Wen, Y.D., Wang, H., Kho, S.H., Rinkiko, S., Sheng, X., Shen, H.M., and Zhu, Y.Z. 2013. Hydrogen Sulfide Protects HUVECs against Hydrogen Peroxide Induced Mitochondrial Dysfunction and Oxidative Stress. *PLoS One* 8:e53147.
- Davidson, S.M. and Duchon, MR. 2007. Endothelial mitochondria: Contributing to vascular function and disease. *Circ Res*.100: 1128-1141.
- Mulvihill, E.E. and Huff, M.W. 2012. Protection from metabolic dysregulation, obesity, and atherosclerosis by citrus flavonoids: activation of hepatic PGC1 α -mediated fatty acid oxidation. *PPAR Research*. 857142.
- Kurowska, E.M., Manthey, J.A., Casaschi, A., and Theriault A.G. 2004. Modulation of HepG2 Cell Net Apolipoprotein B Secretion by the Citrus Polymethoxyflavone, Tangeretin. *Lipids* 39:143-151.
- Yoon J.H., Lim T.G., Lee K.M., Jeon A.J., Kim S.Y., and Lee K.W. 2011. Tangeretin Reduces Ultraviolet B (UVB)-Induced Cyclooxygenase-2 Expression in Mouse Epidermal Cells by Blocking Mitogen-Activated Protein Kinase (MAPK) Activation and Reactive Oxygen Species (ROS) Generation. *J Agric Food Chem*.59:222.
- Chen, Z.T., Chu, H.L., Chyau, C.C., Chu, C.C., and Duh, P.D.2012. Protective effects of sweet orange (*Citrus sinensis*) peel and their bioactive compounds on oxidative stress. *Food Chem*.135:2119.

ikhlas jenie turnitin 1

ORIGINALITY REPORT

23%

SIMILARITY INDEX

18%

INTERNET SOURCES

21%

PUBLICATIONS

7%

STUDENT PAPERS

PRIMARY SOURCES

1	journals.plos.org Internet Source	7%
2	Zong-Tsi Chen, Heuy-Ling Chu, Charng-Cherng Chyau, Chin-Chen Chu, Pin-Der Duh. "Protective effects of sweet orange (Citrus sinensis) peel and their bioactive compounds on oxidative stress", Food Chemistry, 2012 Publication	2%
3	Systems Biology of Free Radicals and Antioxidants, 2014. Publication	2%
4	hyper.ahajournals.org Internet Source	2%
5	Submitted to University of Bristol Student Paper	1%
6	www.tandfonline.com Internet Source	1%
7	e-sciencecentral.org Internet Source	1%

8	academicjournals.org Internet Source	1%
9	jjnpp.com Internet Source	1%
10	worldwidescience.org Internet Source	1%
11	Ya-Dan Wen, Hong Wang, Sok-Hong Kho, Suguro Rinkiko, Xiong Sheng, Han-Ming Shen, Yi-Zhun Zhu. "Hydrogen Sulfide Protects HUVECs against Hydrogen Peroxide Induced Mitochondrial Dysfunction and Oxidative Stress", PLoS ONE, 2013 Publication	1%
12	pubs.sciepub.com Internet Source	1%
13	www.mdpi.com Internet Source	1%
14	Submitted to Universitas Jambi Student Paper	1%
15	Submitted to Higher Education Commission Pakistan Student Paper	1%
16	Y S Hartini, L Nugroho. " The accumulation of two neolignan in the leaves, stems, and flower of red betel (Ruiz & Pav.) ", Journal of Physics:	1%

Conference Series, 2017

Publication

Exclude quotes On

Exclude bibliography On

Exclude matches < 1%